Developmental and hormonal regulation of gene expression of fibrillin-1 (FBN1) and the asprosin receptor, olfactory receptor family 4 subfamily M member 1 (OR4M1), in bovine ovarian cells

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BACKGROUND

- Asprosin is a novel protein encoded by *FBN1* gene and formed when FBN1 is cleaved at C-terminal end by enzyme furin (Duerrschmid et al., 2017).
- Asprosin is correlated with insulin resistance, type II diabetes and PCOS and functions via the G-protein-cAMP-PKA pathway and increase hepatic glucose level (Romere et al., 2016).
- Asprosin receptor, olfactory receptor (Olfr734/OR4M1) has been identified in liver, kidney, and testes (Li et al., 2019).
- HYPOTHESIS: FBN1 mRNA is expressed in ovarian follicular cells and is hormonally regulated.

OBJECTIVES

- To characterize *FBN1, furin* and *OR4M1* mRNA abundance in theca (TC) and granulosa cells (GC) during follicular development and the possible hormone regulators for the expression of *FBN1* mRNA.
- To identify the effects of asprosin on steroid production and proliferation of TC.

MATERIALS AND METHODS

- **Biological Materials.** GC and TC of small (<5mm) and large (>6 mm) follicles were obtained at abattoir. Cells were collected for RNA isolation or cultured with 10% fetal calf serum and after 48 h, treatments were applied in a serum-free medium for 24 h.
- RNA extraction was done using Trizol method.
- Gene expression analysis was made using one-step rt-PCR with 18s as reference gene. Data are presented as $2^{-\Delta\Delta Ct}$.
- **Cell counting and radioimmunoassay (RIA).** Cells were counted using Coulter Counter. Medium from culture plates were collected for double antibody RIA for quantification of progesterone and androstenedione production.
- **Statistical analysis.** Data were analyzed by 2x2 factorial (Exp. 1), 3x2 factorial (Exp. 2), one way ANOVA (Exp. 3) and 2x2 factorial (Exp. 4).

RESULTS

- Experiment 1: FBN1, OR4M1 (Figure 1) and furin (data not shown) mRNA abundance varied with follicle development and cell type.
- Experiment 2: FBN1 mRNA abundance in TC was increased (P < 0.05) by TGFβ1 (Figure 2), FGF2, FGF9 and EGF (data not shown).
- Experiment 3: TGFβ1, WNT3A and FGF9 increased (P < 0.05) FBN1 mRNA abundance in GC and IGF1 decreased (P < 0.05) FBN1 mRNA in GC in vitro (data not shown).
- Experiment 4: Asprosin inhibited (by 10%; P < 0.05) IGF-1 induced TC numbers, but had no effect on progesterone production (P > 0.10). Asprosin increased androstenedione production by 26% in LH treated TC (P < 0.05) (data not shown).

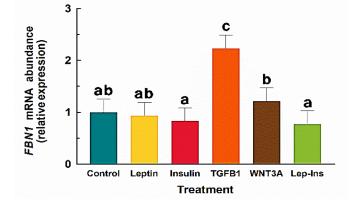
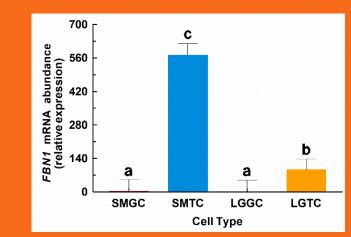


Figure 2: (Exp. 2) TGFβ1 induced *FBN1* mRNA abundance in theca cells. Bars without common letter differ (*P* < 0.05).

CONCLUSION: *FBN1* and *OR4M1* mRNA are developmentally and hormonally regulated in ovarian GC and TC and may perform a paracrine/ autocrine regulation of ovarian function in cattle by regulating cell proliferation and stimulating hormone-induced androgen production.

Literature Cited: Duerrschmid et al., 2017, Nature Med. 23:1444. Li et al., 2019, Cell Metab. 30:319. Romere et al., 2016, Cell 165:566.



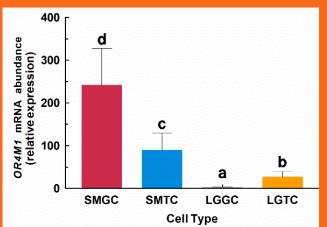


Figure 1 (Exp. 1). FBN1 mRNA abundance is greater in TC than GC (top), whereas, OR4M1 mRNA is greatest in small follicle GC (bottom). SM-small, LG-large, GC-granulosa cells, TC-theca cells. Bars without a common letter differ (P < 0.06).

Asprosin may be acting as a autocrine/ paracrine regulator of ovarian follicular function in cattle.



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