

Background

- Since the gut of young mammals is devoid of microorganisms at birth, its colonization by symbionts and their arrangement into complex communities is a critical aspect of a young host's post-natal development
- Indeed, these microbial communities allow for increased health and nutrition utilization in young calves at the most vulnerable time in their lives.
- A better understanding of how the gut microbiome develops is therefore critical for improving calf health and optimizing their later productivity as mature animals

Objectives

- Determine the gut bacterial composition of 12 neonatal dairy calves using fecal samples
- Identify prominent known and uncharacterized gut bacterial species in growing dairy calves

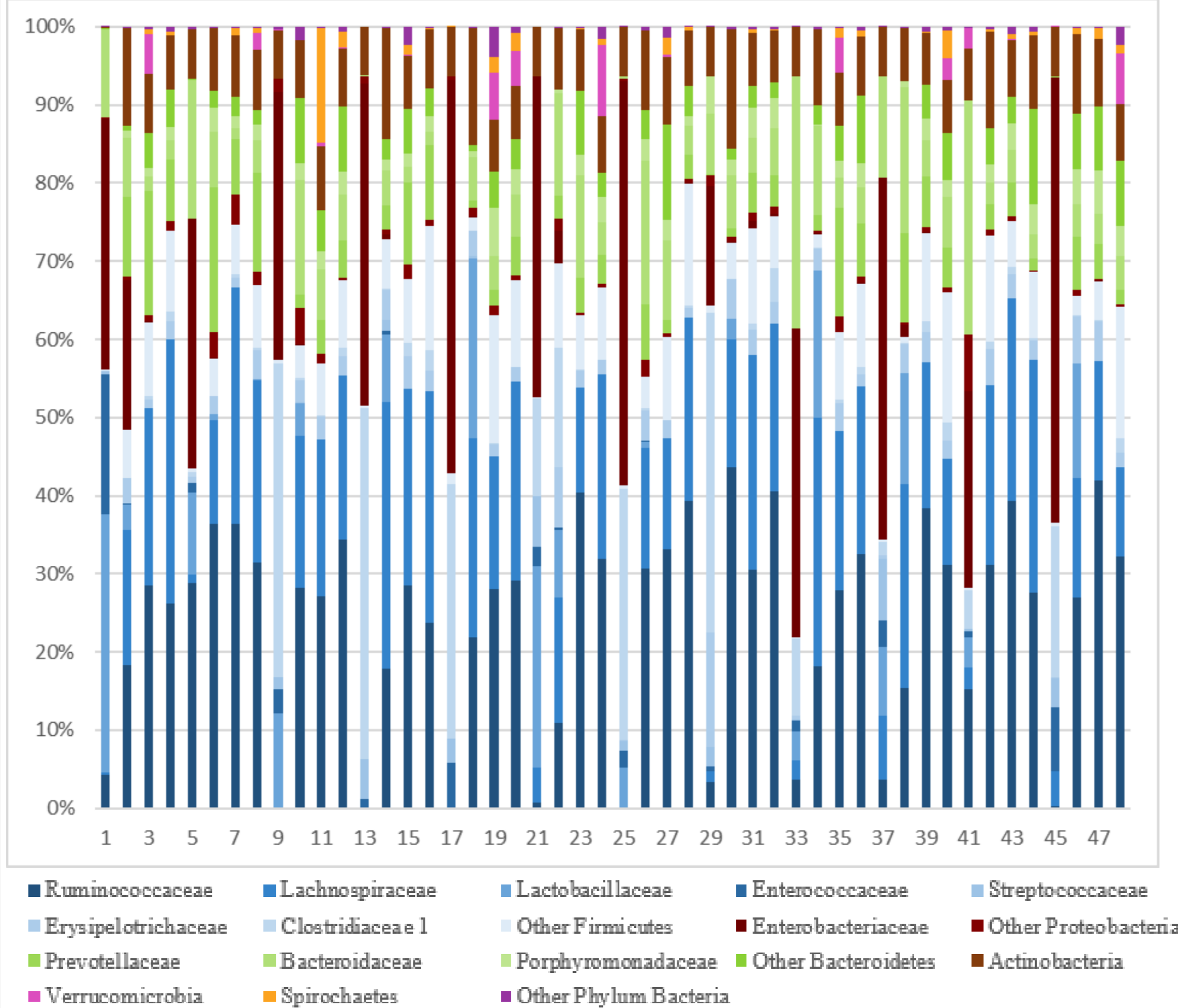
Methods

- Fresh fecal samples were collected from 12 dairy calves. Samples were collected at weeks 0, 4, 8 and 12 after birth.
- Microbial DNA was extracted using a repeated bead beating plus column method.
- 16S rRNA gene sequence diversity was determined by high-throughput sequencing (MiSeq 2X300, Illumina) of PCR-generated amplicons
 - 27F – 519R primers (V1-V3 region)
- Operational Taxonomic Unit (OTU) clustering was performed using custom-written Perl scripts (96% cutoff)
- Taxonomic affiliation was determined using the online tools RDP classifier and BLAST
- Alpha diversity indices and Principal Coordinate Analysis (PCoA) were performed using the open source software MOTHUR (v.1.44.1)
- Kruskal-Wallis and Wilcoxon-pairwise tests were performed in R (Version 3.6.2)

Table 1. Comparative analysis of main taxonomic groups between time points. Different superscripts indicate statistical difference ($P < 0.05$).

Taxonomy	Week 0	Week 4	Week 8	Week 12
Firmicutes				
Clostridiaceae 1	21.30 ^a ± 5.30	1.51 ^b ± 1.34	0.60 ^{bc} ± 0.17	1.35 ^c ± 0.40
Ruminococcaceae	5.31 ^a ± 2.65	26.50 ^b ± 2.92	35.48 ^c ± 1.72	33.78 ^c ± 1.51
Lactobacillaceae	9 ^a ± 3.16	8.84 ^a ± 2.37	< 0.01 ^b	0.01 ^b ± < 0.01
Lachnospiraceae	2.25 ^a ± 0.78	22.43 ^b ± 2.13	22.19 ^b ± 1.68	24.65 ^b ± 1.94
Enterococcaceae	4.13 ^a ± 1.43	0.09 ^b ± 0.05	< 0.01 ^c	< 0.01 ^c
Erysipelotrichaceae	1.48 ^a ± 1.29	4.38 ^b ± 0.48	3.06 ^{bc} ± 0.37	2.42 ^c ± 0.21
Streptococcaceae	2.89 ^a ± 0.82	0.16 ^b ± 0.11	< 0.01 ^c	0 ± 0 ^c
Other Firmicutes	0.53 ± 0.11	5.42 ± 1.16	9.51 ± 1.00	12.25 ± 1.15
Bacteroidetes				
Porphyromonadaceae	0.44 ^a ± 0.43	1.88 ^b ± 0.38	3.15 ^c ± 0.48	2.94 ^c ± 0.28
Bacteroidaceae	9.90 ± 3.65	10.10 ± 1.68	5.39 ± 1.04	4.63 ± 0.45
Prevotellaceae	0.04 ^a ± < 0.01	6.10 ^b ± 1.65	7.10 ^b ± 1.37	5.86 ^b ± 1.01
Other Bacteroidetes	0.02 ± < 0.01	3 ± 0.83	5.86 ± 0.86	5.91 ± 0.98
Proteobacteria				
Enterobacteriaceae	41.22 ^a ± 3.68	2.35 ^b ± 1.69	0.12 ^{bc} ± 0.07	0.04 ^c ± 0.03
Other Proteobacteria	1.08 ± 0.63	1.44 ± 0.40	1.25 ± 0.30	0.76 ± 0.14
Verrucomicrobia				
Verrucomicrobiaceae	0.24 ^a ± 0.23	< 0.01 ^b	1.50 ^c ± 0.72	2.24 ^{ac} ± 0.94
Actinobacteria				
Coriobacteriaceae	6.21 ^a ± 0.56	11.48 ^b ± 0.93	7.93 ^c ± 0.24	7.83 ^c ± 0.22
Spirochaetae				
Spirochaetaceae	< 0.01 ^a	0.12 ^b ± 0.08	2.30 ^c ± 1.23	1.11 ^c ± 0.31
Other Bacteria	0.09 ± 0.04	0.34 ± 0.14	0.93 ± 0.35	0.74 ± 0.20

Figure 1. Taxonomy-based composition analysis of fecal bacterial communities in individual samples collected from neonatal dairy calves.



Results

Table 2. Comparative analysis of α diversity indices between time points. Different superscripts indicate statistical difference ($P < 0.05$).

Index	Week 0	Week 4	Week 8	Week 12
Ace	197.50 ^a ± 33.81	445.20 ^b ± 44.21	770.73 ^c ± 26.40	960.77 ^d ± 52.13
Chao	117.39 ^a ± 16.18	372.50 ^b ± 36.64	682.15 ^c ± 20.45	850.26 ^d ± 34.80
Shannon	1.83 ^a ± 0.06	3.89 ^b ± 0.12	4.76 ^c ± 0.02	5.05 ^d ± 0.05
Simpson	0.27 ^a ± 0.02	0.05 ^b ± 0.01	0.02 ^c ± 0.003	0.02 ^c ± 0.001
Sobs	64.42 ^a ± 4.08	250.33 ^b ± 24.61	451.58 ^c ± 15.31	550 ^d ± 13.98

Table 3. Comparative analysis between time points of the most abundant OTUs. Different superscripts indicate statistical difference ($P < 0.05$).

OTUs	Week 0	Week 4	Week 8	Week 12	Closest valid taxon (id%)
Firmicutes					
JA_17-00190	1.6 ^a ± 4.6	.90 ^b ± .90	< 0.01 ^b	< 0.01 ^b	<i>Cl perfringens</i> (98.73%)
JA_90-20062	1.0 ^a ± 1.0	7.3 ^b ± 2.6	.72 ^c ± 2.2	0.23 ^a ± 0.01	<i>F. prausnitzii</i> (96.57%)
JA_13-22430	3.3 ^a ± 1.8	4.0 ^a ± 1.3	< 0.01 ^b	< 0.01 ^b	<i>Lactobacillus johnsonii</i> (99.63%)
JA_70-18057	< 0.01 ^a	3.1 ^b ± 0.64	1.7 ^b ± 0.39	0.64 ^c ± 0.15	<i>Blautia wexlerae</i> (96.59%)
JA_46-21334	0.76 ^a ± 0.66	4.5 ^b ± 1.22	< 0.01 ^c	< 0.01 ^c	<i>Lactobacillus reuteri</i> (99.26%)
JA_81-39081	4.1 ^a ± 1.81	6.0 ^a ± 0.22	< 0.01 ^b	0 ^b ± 0	<i>Butyrivibrio bullicorcorum</i> (95.19%)
JA_18-34620	0.01 ^a ± < 0.01	4.5 ^b ± 1.7	0.03 ^a ± 0.02	0.01 ^a ± < 0.01	<i>Blautia stercori</i> (96.18%)
JA_16-68527	< 0.01 ^a	0.03 ^a ± 0.03	2.4 ^b ± 0.92	1.7 ^b ± 0.489	<i>Oscillibacter ruminantium</i> (90%)
JA_5-29412	4.0 ^a ± 2.12	0.04 ^a ± 0.027	< 0.01 ^b	< 0.01 ^b	<i>Lactobacillus murinus</i> (100%)
JA_46-37837	< 0.01 ^a	2.76 ^b ± 1.03	0.89 ^b ± 0.25	0.26 ^c ± 0.12	<i>Blautia caecimuris</i> (97.09%)
JA_33-48728	2.95 ^a ± 1.40	< 0.01 ^b	< 0.01 ^c	< 0.01 ^c	<i>Enterococcus lactis</i> (99.36%)
JA_36-17512	< 0.01 ^a	0.04 ^a ± 0.04	1.22 ^b ± 0.40	1.41 ^b ± 0.41	<i>Oscillibacter valericigenes</i> (90.28%)
JA_27-17537	< 0.01 ^a	0.20 ^a ± 0.20	1.00 ^b ± 0.30	1.14 ^b ± 0.27	<i>Oscillibacter ruminantium</i> (89.02%)
JA_84-17601	< 0.01 ^{ab}	0.20 ^a ± 0.20	0.64 ^b ± 0.51	1.4 ^c ± 0.33	<i>Oscillibacter ruminantium</i> (89.21%)
JA_17-04639	1.68 ^a ± 0.69	0.01 ^b ± 0.01	0.09 ^b ± 0.04	< 0.01 ^b	<i>Clostridium paraputrificum</i> (99.80%)
JA_5-10150	1.60 ^a ± 0.62	0.14 ^b ± 0.11	< 0.01 ^c	0 ^c ± 0	<i>Streptococcus macedonicus</i> (99.25%)
Bacteroidetes					
JA_18-16727	< 0.01 ^a	1.7 ^b ± 0.65	4.7 ^c ± 0.86	4.6 ^c ± 0.87	<i>Muribaculum intestinale</i> (89.39%)
JA_35-04049	< 0.01 ^a	3.6 ^b ± 1.3	0.24 ^c ± 0.09	0.47 ^{bc} ± 0.11	<i>Bacteroides caprophilus</i> (96.77%)
JA_7-08334	< 0.01 ^a	0.78 ^b ± 0.51	1.91 ^b ± 0.72	1.07 ^b ± 0.34	<i>Prevotella copri</i> (99.24%)
JA_54-41198	0.03 ^a ± 0.03	0.88 ^b ± 0.33	1.06 ^b ± 0.29	0.84 ^b ± 0.24	<i>Bacteroides uniformis</i> (99.81%)
JA_16-69458	< 0.01 ^a	0.37 ^b ± 0.27	0.94 ^b ± 0.34	1.44 ^b ± 0.35	<i>Prevotella shahii</i> (90.53%)
JA_5-13995	< 0.01 ^a	1.22 ^b ± 0.55	0.22 ^b ± 0.07	0.42 ^b ± 0.20	<i>Prevotella stercora</i> (99.05%)
Proteobacteria					
JA_89-27339	39.33 ^a ± 3.6	2.3 ^b ± 1.7	0.12 ^{bc} ± 0.07	0.05 ^c ± 0.03	<i>Shigella sonnei</i> (99.81%)
Verrucomicrobia					
JA_36-61905	0.01 ^a ± < 0.01	< 0.01 ^b	1.47 ^c ± 0.72	2.23 ^c ± 0.94	<i>Methylococcoides burtonii</i> (91.43%)
Actinobacteria					
JA_6-16768	< 0.01 ^a	3.34 ^b ± 0.91	< 0.01 ^a	< 0.01 ^a	<i>Collinsella aerofaciens</i> (98.38%)

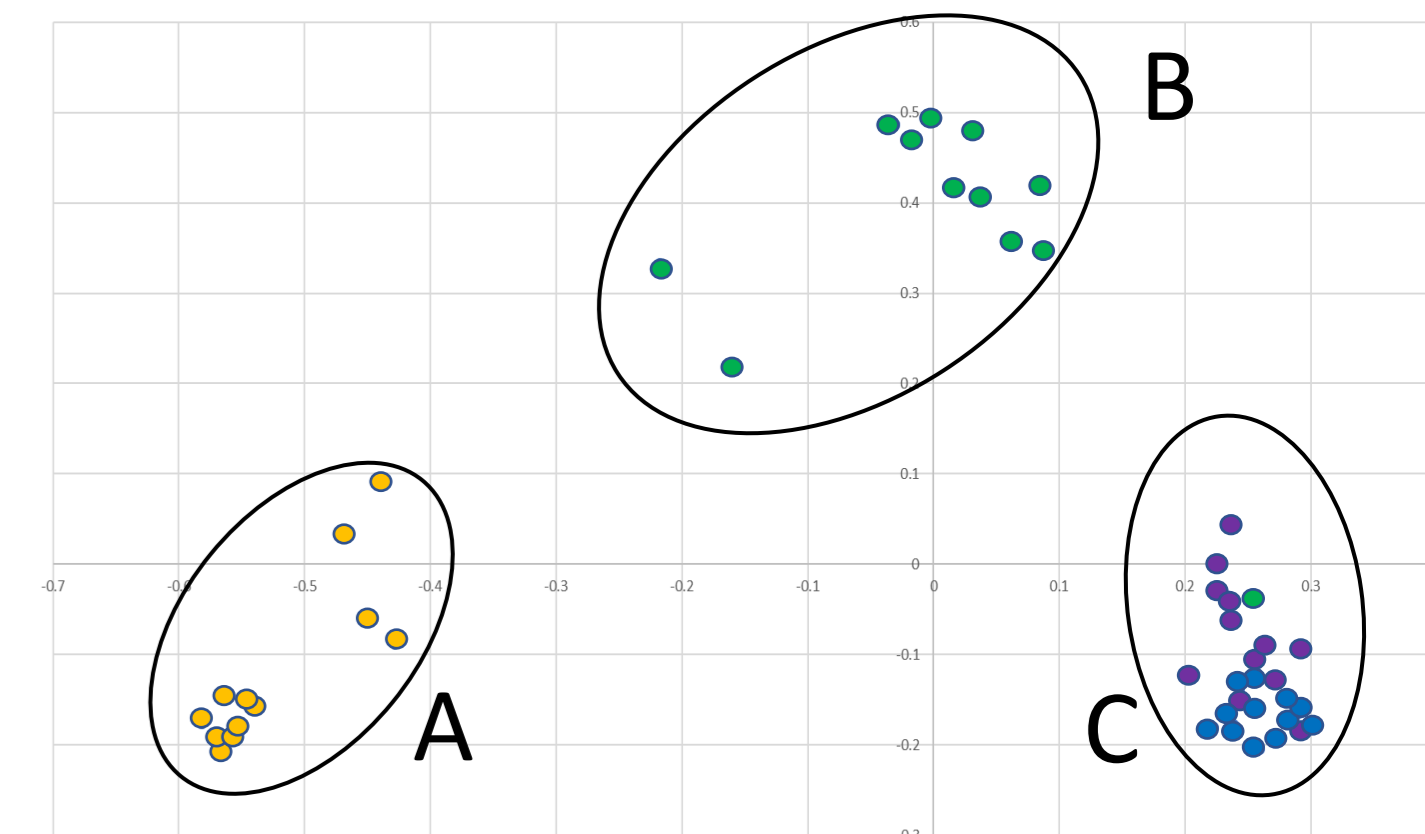


Figure 2. β diversity analysis by PCoA. Each sample is represented by a circle, with distance between points indicating the relative similarity in bacterial community structure between them. Samples were found to form three main groups (A, B, C), each clustered in a fashion consistent with their time of collection. Cluster A - Week 0 (yellow) Cluster B - Week 4 (green) Cluster C - Week 8 (purple) ;and Week 12 (blue)

Figure 1 Legend.

Samples (1-48) with corresponding week
 Week 0 - 1,5,9,13,17,21, 25, 29,33,37,41,45
 Week 4 - 2,6,10,14,18,22,26,30,34,38,42,46
 Week 8 - 3,7,11,15,19,23,27,31,35,39,43,47
 Week 12- 4,8,12,16,20,24, 28,32,36,40,44,48

Conclusions

- A gradual increase in species diversity was observed during the trial, with 3 distinctive bacterial community compositions observed at week 0, week 4 and weeks 8-12, respectively.
- Changes in composition were observed at both the levels of taxonomy and OTUs.
- The respective abundance of 25 distinct OTUs, representing a combination of known and uncharacterized bacterial species, was found to fluctuate during the time period of study.
- Together, these results are indicative of microbial succession taking place in the gut of neonatal dairy calves, with the period of week 0 to week 8 representing the time when gut bacterial communities are most fluid in composition.

Acknowledgements

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