# Whole Genome Sequencing Analysis of *Klebsiella pneumoniae* Isolates Reveals Diversity in Genetic Antibiotic Resistance Patterns

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CLUSTERS								
al 1	Unit A		CG-258					
	Unit B						CC-20	F
	Unit C	CG-15	CG-15	CG-258	CC-20	CC-20	CC-20	(
	Unit D	CG-15		CG-258	CC-20		CG-15	Γ
it.	Unit E	CG-258		CG-258	CC-20			
ġ,				CG-258				
HOS	Unit F	CG-258	CG-15		CC-20	CC-20		
			CG-15					
			CG-15					
	Unit G					CC-20	CG-15	Г
2	Unit H	CG-258	CG-258	CC-20	CC-20			
		CG-258		CC-20	CC-20			
al				CC-20	CC-20			
lospitá	Unit I	CG-258					CG-258	
	Unit J	CG-258						
	Unit K		CG-15					
	Unit L	CG-258	CG-15	CC-20		CG-258	CG-258	
н			CG-15					
			CG-15					

**Table 1.** Clusters arranged by hospital and by unit. No predominant or localized cluster identified. Arranged for visual clarity; does not reflect actual geographic room locations.

STRAINS								
Hospital 1	Unit A		ST-258					
	Unit B						ST-1243	
	Unit C	ST-14	ST-15	ST-348	ST-29	ST-280	ST-584	N/A
	Unit D	ST-14		ST-348	ST-29		ST-551	
	Unit E	ST-11		ST-348	ST-29			
				ST-348				
	Unit F	ST-11	ST-15		ST-29	ST-20		
			ST-15					
			ST-15					
	Unit G					ST-20	ST-107	
2	Unit H	ST-11	ST-258	ST-16	ST-1694			
Hospital		ST-11		ST-16	ST-1694			
				ST-16	ST-1694			
	Unit I	ST-11					ST-2670	
	Unit J	ST-11						
	Unit K		ST-15					
	Unit L	ST-11	ST-15	ST-16		ST-834	ST-54	
			ST-15					
			ST-15					

Table 2. Strains arranged by hospital and by unit. Four hospital-specific and one unitspecific strain identified by MST analysis. Arranged for visual clarity ; does not reflect actual geographic room locations.







# BACKGROUND

G-258	

### Klebsiella pneumoniae is among the leading causes of healthcare-associated infections (HAI). Multidrug-resistant (MDR) Klebsiella variants are difficult to treat and have been reported with increasing frequency in hospitals. Under the hypothesis that nosocomial strains should bear genetic similarities if they are breeding and spreading throughout a hospital or unit, we sought to compare genetic similarities/differences of hospital-acquired Klebsiella pneumoniae isolates using whole genome multi-locus sequence typing (wg-MLST) analysis, and then compare the antibiotic resistance genetic patterns against their epidemiologic typing.

## METH(

- We attempted to find hospital and/or unit-specific nosocomial K. pneumo hospitals in the Detroit, Michigan area. Samples were collected after 48 h total of 46 clinical bacterial HAI isolates were collected between 2017-20
- We extracted genomic DNA using the QIAamp DNA Micro Kit (Qiagen, Hil (Illumina, San Diego, CA) to build libraries from 200 ng of DNA and genera and instrument (Illumina, San Diego, CA).
- We de novo assembled the genome using the SPAdes (version 3.7.1) asser sequence type (ST) was then assigned for each isolate.
- We then compared each isolate the international ResFinder Database (ver genes for each isolate.
- We then analyzed for patterns against geographic hospital and unit, as we

### RESUI

There was significant genetic diversity among the isolates, and

### **CLUSTER & STRAIN ANALYSIS**

- Isolates belonged to one of three distinct phylogenetic clusters: G-258 | CG-15 | CG-20.
- However, no cluster was hospital or unit specific, and each cluster contained multiple strains.
- MLST analysis revealed 17 unique strains. Seven strains had genetically unique resistance genes detected in more than one isolate, but only five of these seven were hospital-specific.
  - ST-14 did not have any unique or specific ABX resistance genes **ST-16** was unique for several *bla-SHV* (beta-lactam) genes (26, 78, 98, 145, 179, 194, 199). It represents 4 of the 22 samples from Hospital 2 and was found across two different units.
  - ST-29 did not have any unique or specific ABX resistance genes
  - **ST-348** was unique for the *bla-SHV-81* gene ST-1694, the only unit-specific strain, lacked any unique or
  - strain-specific ABX resistance genes.

# CONCLU

Genetic analysis of antibiotic resistance patterns and wg-MLST serotypes revealed significant heterogeneity among HAI K. pneumoniae isolates, much more than was initially expected. The low prevalence of specific genes and absence of geographic clustering indicates no common source of transmission for either hospital. Although K. pneumoniae is a very common nosocomial pathogen, etiologic analysis suggests diverse community strains (e.g. gut colonization) may actually be responsible for previously-designated HAI.







		100%
DDS		10070
oniae strains by sourcing from two geographically disparate tertiary care nours of admission to select for HAI etiologies not present on arrival. A .019 for analysis. Iden, Germany). We used the Nextera® DNA Flex Library Prep Kit ated paired-end reads (2 × 151 bp) using the Illumina NextSeq reagent kit		96%
mbler on the Bionumerics calculation engine, and unique MLST Pasteur		74%
rsion 3.2) to search for known chromosomal mutations and resistance		78%
ell as within each ST.		78%
		7070
ር.ጥር		
nd no predominant strain or gene pattern was identified.		
<u>GENE ANALYSIS</u>		100%
75 unique resistance genes detected among all isolates. Only 8 hospital-specific genes detected in more than one isolate, and most with very low prevalence:		
<u>Hospital 1</u> : qnrB1 (43%), blaSHV-110 (17%), blaSHV-1 (9%)	_	250/
Hospital 2: <i>blaCTX-M-3</i> (23%), <i>ac(6')-Ib3</i> (18%),	-	35%
blaSHV-36 (9%), blaSHV-80 (9%), blaSHV-178 (9%)		45%
Most prominent besolved 1 specific gaps $(aw R1)$ per specifically	-	0%
found in all 3 clusters, 4 of 6 units, and 5 of 12 strains.		0%
No colistin resistance genes were detected.		0%
Carbapenemase resistance genes were detected in two isolates.	-	0%
Comparisons of phenotypic expression against gene detection show		0%
significant unterences when trying to establish hosocomial patterns.	,	Tahlo 2 I
		expected
SION	]	prevalence clinical sig



ANTIBIOTIC RESISTANCE							
Phenotypic Prevalence			Genotypic Prevalence				
Hospital 1 (N=23)	Hospital 2 (N=22)	Drug Class	Resistance Gene	Hospital 1 (N=23)	Hospital 2 (N=22)		
1000/	1000/		fosA	100%	77%		
100%	100%	FUSFUINTCIN	fosA5	0%	23%		
	100%	QUINOLONE	oqxA	100%	100%		
100%			oqxB	100%	100%		
			qnrB1	43%	0%		
		AG / FQ	aac(6')-Ib-cr	74%	50%		
			aadA2	22%	55%		
			aac(3)-IIa	52%	14%		
96%	86%	AMINOGLYCOSIDE	aph(3'')-Ib	52%	14%		
			aph(6)-Id	52%	14%		
			aph(3')-Ia	22%	32%		
74%	36%	PHENICOL	catB3	74%	32%		
700/	770/	TDINAETUODDINA	dfrA12	22%	55%		
/ð70	//%	I KIIVIE I HOPKIIVI	dfrA14	48%	27%		
	64%	SULPHONAMIDE	sul1	26%	64%		
78%			sul2	48%	14%		
			sul3	9%	18%		
	100%		<i>blaCTX-M-15</i> [ESBL]	87%	64%		
			<i>blaOXA-1</i> [ESBL]	74%	32%		
			<i>blaTEM-1B</i> [ESBL]	57%	50%		
		β-LACTAM	blaSHV-28	17%	18%		
100%			blaSHV-106	17%	18%		
TOO \0			blaSHV-182	13%	23%		
			<i>blaCTX-M-3</i> [ESBL]	0%	23%		
			blaFOX-5 [Amp-C]	4%	0%		
			blaKPC-2	0%	5%		
			blaKPC-3	0%	5%		
35%	64%	MACROLIDE	mph(A)	30%	64%		
43%	9%	TETRACYCLINE	tet(A)	30%	5%		
13%	18%	RIFAMPICIN	ARR-3	13%	18%		
0%	0%	COLISTIN					
0%	0%	FUSIDIC ACID					
0%	0%	GLYCOPEPTIDE					
0%	0%	NITROIMIDAZOLE					
0%	0%	OXAZOLIDINONE					

Hospital-specific prevalence for specific ABX resistance genes as well as phenotypic drug class expression on traditional susceptibility testing. Lowce genes < 18% are omitted for visual clarity (except for KPC and Amp-C for gnificance).

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