Temporal Dynamics of the Plasma Microbiome in Recipients at Early Post-liver Transplantation IDWeek 2020 Submission ID: 902309

Toshihiko Okumura 1, Kazuhiro Horiba 1.2.3, Hideya Kamei 4, Suguru Takeuchi 1, Takako Suzuki 1, Yuka Torii 1, Jun-ichi Kawada 1, Yoshiyuki Takahashi 1, Yasuhiro Ogura 4, Tomoo Ogi 2.3, and Yoshinori Ito 1 ¹ Department of Pediatrics, Nagoya University Graduate School of Medicine, Nagoya, Japan ² Department of Genetics, Research Institute of Environmental Medicine, Nagoya University, Nagoya, Japan ³ Department of Human Genetics and Molecular Biology, Nagoya University Graduate School of Medicine, Nagoya, Japan ⁴ Department of Transplantation Surgery, Nagoya University Hospital, Nagoya, Japan

Contact Information Nagoya university Graduate School of Medicine 65 Tsurumai-cho, Showa-ku, Nagoya 466-8550 E-mail: t-okumura@med.nagoya-u.ac.jp TEL/FAX: +81-52-744-2294/2974

C

4.5

p=0.03

Introduction

Immunosuppression during liver transplantation (LT) enables the prevention and treatment of organ rejection, but poses a risk for severe infectious diseases. Immune modulation and antimicrobials affect the blood microbiome. Thus, determining the impact of immunosuppression and antimicrobials on the microbiome may be important to understand immunocompetence, predict clinical adverse events after LT such as acute cellular rejection (ACR), and treat infectious diseases. In this study, we characterized the blood microbiome of LT recipients using next-generation sequencing (NGS). Moreover, NGS was evaluated as a detection tool of causative pathogens.

Conclusions

- Anelloviridae and Enterobacteriaceae abundance dynamically changed in the early phase of post-transplantation
- The NMDS analysis revealed that antimicrobials are associated with the microbiome composition.
- Enterobacteriaceae abundance and high blood microbiome diversity may be an indicator for the development of ACR after LT.
- NGS is useful as a comprehensive diagnostic tool for pathogens in LT recipients.

Materials and Methods

Patients and Samples Plasma samples were collected according to the criteria below; Fifty-one LT recipients 1) within a week after LT (46 samples) 2) 4 ± 1 weeks after LT (28 samples) Nagoya University Hospital 2016 to 2018 3) 8 ± 1 weeks after LT (29 samples) 4) within 2 days after a positive blood culture (16 samples)

Table. Characteristics of the patients

	Total (n = 51)		n = 51
No. patients (pediatric/adult)	13/38	Underlying disease	
Age at LT [median (range)], pediatric adult	12 m (5–50 m) 50.5 y (18–66 y)	Biliary atresia Primary sclerosing cholangitis Primary biliary cholangitis	10 (19.6%) 9 (17.6%) 6 (11.8%) 6 (11.8%)
Sex (male/female)	19/32	Fulminant hepatic failure	
Living donor/brain death donor	38/13	Others	20 (39.2%)
ABO-incompatible	8 (15.7%)	ACR	19 (37.3%)

Sample Preparation

- Extraction DNA from 140 µL of plasma
- Nextera XT Library Preparation Kit (illumina)
- Library quality assessment with Bioanalyser (Agilent) and Droplet digital PCR
- Sequencing with NGS
- Hiseq (illumina), 150bp Paired-end sequencing
- Target output was 10,000,000 reads/sample
- Detection of Microorganism Genomes
- Metagenomic analyses were performed through the custom-made analysis pipeline PATHDET (PATHDET v1.0, https://pathdet.hgc.ip/)

PATHDET

- Relative abundance
- Simpson's diversity index and Shannon's diversity index





A100%

Figure 1.

Results

(A) Change in the relative abundance of each microorganism at the family level (B and C) Change in plasma microbiome diversity at the genus level

- Enterobacteriaceae (§) abundance decreased in samples collected at week 4 and 8 posttransplantation in comparison with samples collected at week 1 post-transplantation.
- Simpson's diversity index of samples collected at week 1 post-transplantation was significantly higher than that of samples collected at week 8 post-transplantation (0.85 \pm 0.09 vs. 0.76 \pm 0.23, respectively, p = 0.03).



Non-metric multidimensional scaling (NMDS) representation of plasma samples at each sampling point with envfit correlations

Arrows represent envfit correlations for (A) time from LT and (B) antimicrobials.

- The NMDS plot of the microbiome showed similarity of the microbiome among samples at the same time point and between samples with and without antimicrobials.
- Antifungals and antivirals were also associated with the microbiome (data not shown).

0.95 ŝ Corynebacteriaceae Streptococcaceae 0.9 ළී 3.5 Micrococcaceae 0.85 Methylobacteriaceae 0.8 Sphingomonadaceae ≥ 0.75 2.5 Burkholderiaceae Moraxellaceae 0.7 Xanthomonadaceae* 0.65 Herpesviridae 1.5 Staphylococcaceae 0.6 Pseudomonadaceae 0.55 Propionibacteriaceae 0.5 0.5 Nocardiaceae Anelloviridae 0.45 ACR non-ACR ACR non-ACR Comamonadaceae Enterobacteriaceae

Others

Figure 3.

(A) Comparison of the plasma microbiome at the family level in patients with and without ACR (B and C) Comparison of plasma microbiome diversity at the genus level in patients with and without ACR

- Enterobacteriaceae (†) abundance was significantly decreased in the ACR group in comparison with the non-ACR group (p = 0.045)
- Simpson's diversity index in the ACR group were significantly higher than those in the non-ACR group (0.89 \pm 0.04 vs. 0.84 \pm 0.10, respectively, p = 0.03).

No	Diagnosis	Bacteria Isolated in Blood Cult	ure	F	Relative A	bundance	3	
16	CRBSI	Staphylococcus aureus	_	_	_			
20	FUO	Corynebacterium striatum						
54	CRBSI	MRSA						
62	cholangitis	Escherichia coli						
67	liver abscess	Enterococcus faecalis						
83	CRBSI	Staphylococcus aureus						
97	CRBSI	Enterobacter cloacae complex						
99	peritonitis	Acinetobacter baumannii			100.00			
101	peritonitis	Stenotrophomonas maltophilia	1					
105	peritonitis	Enterococcus faecium						
106	FUO	Haemophilus influenzae						
107	hepatic biloma	Enterococcus faecium						100 B.
108	hepatic biloma	Enterococcus faecium						
109	hepatic biloma	Enterococcus faecium	_					
			0%	2004	10%	60%	0004	1000
	-		0 / 0	2070	TU /0	00 /0	0070	100

Figure 4.

Relative abundance of microorganisms at the species level in plasma from patients with positive blood cultures

The same bacterial species isolated in blood culture were identified by NGS in 14 of 16 samples, and predominant pathogens in blood culture were identified in 8 samples.

CRBSI, catheter-related bloodstream infection; FUO, fever of undetermined origin; MRSA, methicillinresistant Staphylococcus aureus.